

## **Comparison of APSIM and DNDC when predicting N<sub>2</sub>O emissions from urine patches**

**Iris Vogeler<sup>\*1</sup>, Rogerio Cichota<sup>2</sup> and Donna Giltrap<sup>3</sup>**

<sup>1</sup>Aarhus University, Denmark

<sup>2</sup>Plant and Food Research, New Zealand

<sup>3</sup>Landcare Research, New Zealand

Corresponding author: iris.vogeler@agro.au.dk

**Keywords:** APSIM, DNDC, denitrification, nitrification, nitrous oxide, urine patches

### **Extended Abstract**

Agricultural greenhouse gas (GHG) emissions, including nitrous oxide (N<sub>2</sub>O), are a major contributor to total GHG emissions, and animal excreta deposited onto pastures are a main source of this. To reduce N<sub>2</sub>O emissions better understanding of the factors driving emissions and evaluation of mitigation strategies is required. Computer simulation models can provide an effective tool for these. We compared two process-based models simulating C- and N-cycling in agricultural soils: APSIM<sup>1</sup> and NZ-DNDC<sup>2</sup>, an adapted to New Zealand grazed pasture conditions. The aim was to identify primary differences in simulated nitrogen transformation rates in soils under varying environmental conditions including temperature (T), soil water content (θ), soil organic carbon (SOC), pH, and NH<sub>4</sub> and NO<sub>3</sub> concentration.

### **Model descriptions**

APSIM is a framework of biophysical modules that simulate biological and physical processes in farming systems. The SoilN module simulates the dynamics of N and C on a daily time-step in soil layers, with C and N mineralisation, C and N immobilisation, N nitrification, N denitrification and nitrate and ammonium adsorption and movement explicitly described in each layer. These N processes are controlled by soil water content, temperature and pH and water flow which are simulated within either the SoilWat (tipping bucket model) or SWIM (based on Richards equation) modules.

DNDC consists of several sub-models for simulating thermal-hydraulic flows, plant growth, aerobic decomposition, fermentation and denitrification. The model usually operates on a daily time-step, except following a rainfall event where denitrification is calculated on an hourly time-step. To allow nitrification and denitrification to occur simultaneously in aerobic or anaerobic microsites, a dynamic 'anaerobic balloon' is used<sup>3</sup>. Substrates such as carbon, NH<sub>4</sub> and NO<sub>3</sub> are split into aerobic and anaerobic soil micro-sites.

## Methods

Firstly, to compare N transformations from APSIM and DNDC, simulations were set up with simplified conditions (no plant cover, constant temperature, uniform soil properties with depth). In separate simulations environmental conditions were altered: the initial N concentration (100 or 500 kg NH<sub>4</sub>/ha, or 100 kg NO<sub>3</sub>),  $\theta$  of 0.3, 0.45, 0.55 m<sup>3</sup> m<sup>-3</sup>, soil T of 10, 15 and 30°C, SOC of 3 and 6%, pH of 6 and 8, and +/- rainfall of either 20 mm on day 1, or at 5 mm/day).

APSIM and DNDC were then compared to a series of field measurements of soil mineral N (NH<sub>4</sub> and NO<sub>3</sub>) and N<sub>2</sub>O emissions following urine application, with different soil types, different climates and urine application timings<sup>4</sup>.

## Results

Simulations under the simplified conditions revealed that soil T had a larger influence on the nitrification rate in APSIM than NZ-DNDC, while NH<sub>4</sub> concentration and soil water content had more influence on the nitrification rate in NZ-DNDC than in APSIM. NZ-DNDC showed a higher sensitivity of denitrification rate to T and organic carbon than APSIM<sup>5</sup>.

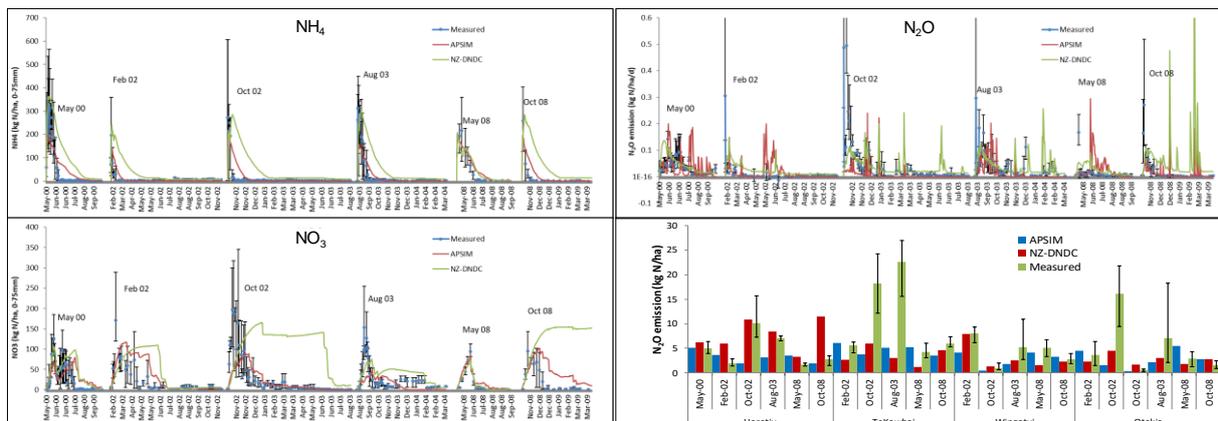


Figure 1. APSIM and NZ-DNDC model results compared with measured values of NH<sub>4</sub>, NO<sub>3</sub>, and N<sub>2</sub>O following urine application for six trials on the Horotiu soil, New Zealand. Also shown are the measured and predicted cumulative emissions.

The two models predict N transformation from the urine patches quite differently, with APSIM in general predicting faster nitrification, an example is shown for the time series of the Horotiu soil, a well-drained allophanic soil (Fig.1). The daily measurements of N<sub>2</sub>O emissions (replicates) show high spatial variability, indicating the sensitivity of N<sub>2</sub>O emissions to variability in soil properties over short distances. Both models frequently produced results outside the measured range, with both over- and underestimation. Total N<sub>2</sub>O emissions also

show mixed results by the two models, with some measurements predicted well by both models, as well as over- or underpredictions. Statistical analysis suggests that APSIM is better at predicting trends between soil types, whereas NZ-DNDC is better at predicting seasonal differences in N<sub>2</sub>O emissions. In New Zealand a specific emission factor (EF3) of 1% for deposited urine, is employed within the national agricultural greenhouse gas inventory<sup>6</sup>. The two process-based models performed less well than the compared to when calculating cumulative N<sub>2</sub>O emissions based on the EF3 method.

An initial sensitivity analysis, in which various model parameters were varied, indicated, that while changing some of the default model parameters improved the agreement in some cases (e.g. for APSIM when denitrification rate was decreased or the fraction of nitrified N emitted as N<sub>2</sub>O was increased, and for DNDC when microbial activity was decreased or volatilization increased), so far none of the model parameters investigated could produce emissions that agreed reasonably with all datasets. A sensitivity analysis, which includes more parameters and model functions, as well as changing various parameters simultaneously, is needed.

One reason for the difficulty in modeling N<sub>2</sub>O emissions is that they are small compared to other N fluxes (e.g. plant uptake, leaching, NH<sub>3</sub> volatilisation). So, simulating these other N transformation processes with low error is required to accurately simulate N<sub>2</sub>O emissions. There have been some subsequent improvements to the simulation of NH<sub>3</sub> volatilisation in NZ-DNDC<sup>7</sup>, which have not been tested in these comparisons.

## Conclusions

This paper presents a comparison of the two different models, APSIM and DNDC to simulate nitrogen transformation rates, including nitrification, denitrification and N<sub>2</sub>O emissions in soils. The two models predicted showed quite different responses to environmental factors. It should be stressed that for both models only default values for the many model parameters were used. Fine tuning of model parameter values based on measurements and improved understanding is likely to increase the prediction capability of the models, and maybe also the agreement between the two models.

**Acknowledgment:** The authors would like to acknowledge funding for this work from the New Zealand

Agricultural Greenhouse Gas Research Centre (NZAGRC) under ‘Integrated Systems’ and Ministry for Primary Industry (formerly MAF) under ‘Sustainable Land Management Mitigation and Adaptation to Climate Change’.

**Accreditation:** This paper was given at the workshop on Climate Change, Reactive Nitrogen, Food Security and Sustainable Agriculture which took place in Garmisch-Partenkirchen, Germany, on 15-16 April 2019, and which was sponsored by the OECD Co-operative Research Programme: Biological Resource Management for Sustainable Agricultural Systems. Financial support was provided through the ‘Global N<sub>2</sub>O Database’ project from Colorado State University and through

the ‘N-tool precise’ project from Aarhus University.

### **OECD disclaimer**

The opinions expressed and arguments employed in this paper are the sole responsibility of the authors and do not necessarily reflect those of the OECD or of the governments of its Member countries.

### **References:**

- [1] Holzworth DP, Huth NI et al.: **APSIM - Evolution towards a new generation of agricultural systems simulation**. *Environ Mod & Softw*, 2014:**62**, 327-350.
- [2] Saggarr S, Giltrap DL, Li C, Tate, KR: **Modelling nitrous oxide emissions from grazed grasslands in New Zealand**. *Agri Eco Environ*, 2007:**119**: 205-216.
- [3] Li C, Aber J, Stange F, Butterbach-Bahl K, Papen H: **A process-oriented model of N<sub>2</sub>O and NO emissions from forest soils: 1. Model development**. *J Geophys Res D*, 2000: **105**, 4369-4384.
- [4] Giltrap D, Vogeler I, Cichota R, van der Weerden T, DeKlein, C: **Comparison and validation of the APSIM and NZ-DNDC models with measurements from urine patches**. *NZ J Agri Res*, 2015:**58**,131-155.
- [5] Vogeler I, Giltrap, D, Cichota R. **Comparison of APSIM and DNDC to simulate nitrogen transformations and N<sub>2</sub>O emissions**. *Sci Tot Environ*, 2013: **465**, 147-155.
- [6] Ministry for the Environment 2013. **New Zealand’s Greenhouse Gas Inventory 1990–2011**. Wellington, Ministry for the Environment. <http://www.mfe.govt.nz/publications/climate-change-environmental-reporting/new-zealands-greenhouse-gas-inventory-1990%E2%80%932011>
- [7] Giltrap D, Saggarr S, Rodriguez J, Bishop P 2017. **Modelling NH<sub>3</sub> volatilisation within a urine patch using NZ-DNDC**. *Nutr Cycl Agroecosys* 108:267-277